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Note

NEW ANTHRAQUINONES FROM *GLADIOLUS GANDAVENSIS*

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Two new anthraquinones, methyl 8-hydroxy-3-methoxy-6,7-methylenedioxy-1-methylanthraquinone-2-carboxylate (gandavensin A, **1**) and methyl 8-hydroxy-3,6,7-trimethoxy-1-methylanthraquinone-2-carboxylate (gandavensin B, **2**), have been isolated from the light petroleum extract of the subterranean corm of *Gladiolus gandavensis* Van Houtt., along with methyl 8-hydroxy-3,6-dimethoxy-1-methylanthraquinone-2-carboxylate (**3**), methyl *trans-p*-methoxycinnamate (**4**), 5,7-dimethoxy-2-methylchromone (**5**), and 5-hydroxy-2-hydroxymethyl-7-methoxychromone (**6**). Their structures were elucidated on the basis of spectral data.

Keywords: *Gladiolus gandavensis*; Anthraquinone; Gandavensin A; Gandavensin B

INTRODUCTION

The genus *Gladiolus* (Iridaceae) contains about 250 species world-wide, one of which (*G. gandavensis* Van Houtt.) is found in China [1]. Phytochemical studies on this genus have revealed the occurrence of anthraquinones [2], alkaloids [3], anthocyanidins [4] and flavonols [5]. *G. gandavensis* Van Houtt, a famous ornamental flower plant, is widely cultivated in China [1], and its subterranean corm has been used in Chinese traditional medicine to treat fractures, pharyngitis, parotitis and lymphnoditis due to its properties of detoxification, detumescence and anodyne [6]. However, no chemical study on this plant had been carried out. In this investigation, from the subterranean rhizomes of *G. gandavensis* Van Houtt, two new anthraquinones, methyl 8-hydroxy-3-methoxy-6,7-methylenedioxy-1-methylanthraquinone-2-carboxylate (gandavensin A, **1**) and methyl 8-hydroxy-3,6,7-trimethoxy-1-methylanthraquinone-2-carboxylate (gandavensin B, **2**), were isolated. In addition, four known compounds were isolated, methyl 8-hydroxy-3,6-dimethoxy-1-methylanthraquinone-2-carboxylate (**3**) [7], methyl *trans-p*-methoxycinnamate (**4**) [8], 5,7-dimethoxy-2-methylchromone (**5**) [9], and 5-hydroxy-2-hydroxymethyl-7-methoxychromone (**6**) [10]. Compound **3** is a new natural product. Their structures were determined predominantly *via* spectral data.

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RESULTS AND DISCUSSION

Compound **1** was obtained as orange needles. The molecular formula $C_{19}H_{14}O_8$ was provided by the molecular ion peak at m/z 370.0678 in the HR-EIMS spectrum. The UV–Vis spectrum with maxima absorption at λ_{max} 213, 287, 319, 403 nm, the IR absorption at ν_{max} 1579 (chelated quinone ketone), 1644 cm^{-1} (unchelated quinone ketone), and the generation of a pink colour with methanolic magnesium acetate suggested **1** is an α -hydroxyanthraquinone [11]. The ^1H NMR signal at δ 12.54 and the IR absorption at 3371 cm^{-1} indicated the presence of a chelated hydroxy group. From the ^1H NMR spectrum, two isolated aromatic protons (δ 7.57, 7.33), one methyl group (δ 2.74) located at C-1 due to the deshielding by the neighbouring carbonyl group [2], two methoxyls (δ 3.89, 3.96) and one methylenedioxy group (δ 6.27) were recognized. From the HMBC correlations of the two aromatic protons (δ 7.57, 7.33) with C-10 (δ 182.0), the ^1H NMR at δ 7.57 and 7.33 could be assigned to H-4 and H-5, respectively. The carbon C-7 (δ 139.0) correlated with H-5 (δ 7.33), methylenedioxy proton (δ 6.27) and hydroxyl proton (δ 12.54); the methylenedioxy group could be located at C-6 and C-7, the hydroxyl at C-8. The protons of 1-Me and H-4 correlated with the C atom at δ 130.3 (C-2) and 122.2 (C-9a) in the HMBC experiment, thus the ^{13}C NMR signal at δ 158.9 could only be assigned to C-3. The methoxy group at C-2 was located based on the HMBC cross signal δ 158.9–3.96 (3H, s). The IR absorption at 1738 cm^{-1} , the ^{13}C NMR signal at δ 167.8, as well as the ^1H NMR signal for methoxy group at δ 3.89 suggested the presence of methoxycarbonyl, which could only be located at C-2. Consequently, compound **1** was determined to be methyl 8-hydroxy-3-methoxy-6,7-methylenedioxy-1-methylanthraquinone-2-carboxylate (gandavensin A).

Compound **2** was also obtained as orange needles. The molecular ion peak at m/z 386.1048 in the HR-EIMS spectrum gave the molecular formula $C_{20}H_{18}O_8$. The visualization of **2** with methanolic magnesium acetate, the UV–Vis absorption at λ_{max} 204, 283, 316 and 418 nm and the IR absorptions at ν_{max} 1627, 1668 cm^{-1} (quinone ketones) revealed an anthraquinone skeleton. From the ^1H NMR spectrum, three methoxyl groups (δ 4.01, 4.02, 4.03), one methyl group (δ 2.72), a chelated hydroxy group (δ 13.08) and a methoxycarbonyl (δ 3.98) were recognized. The assignments of these groups were confirmed by HMBC measurements (Table II below). Thus, compound **2** was identified as methyl 8-hydroxy-3,6,7-trimethoxy-1-methylanthraquinone-2-carboxylate (gandavensin B).

1-Methylanthraquinones have been obtained from seven plants, *viz.* *Rhamnus fallax* (Rhamnaceae) [12], *Aloe saponaria* (Liliaceae) [13], *Eleutherine americana* (Iridaceae) [14], *Rheum sp.* (Polygonaceae) [15], *Gladiolus segetum* (Iridaceae) [2], *Crocus sativus* (Iridaceae) [16] and *Araliorhamnus vaginata* (Rhamnaceae) [17]. As opposed to the plant kingdom, the presence of side chain at C-1 is a common feature of anthraquinones in animals [17]. *G. gandavensis*, therefore, appears to be another source for synthesizing such compounds. 1-Methylanthraquinones may be the significant taxonomic feature of *G. gandavensis* Van Houtt (Fig. 1).

EXPERIMENTAL

General Experimental Procedures

Melting points were determined on an XRC-1 micro-melting point apparatus and are uncorrected. NMR spectra were recorded on a Varian ^{unity}Inova-400 spectrometer, the chemical shifts (δ) are given in ppm (TMS as internal standard). EIMS (70 eV) and HR-EIMS were obtained on a VG AutoSpec-3000 mass spectrometer. IR and UV spectra were carried

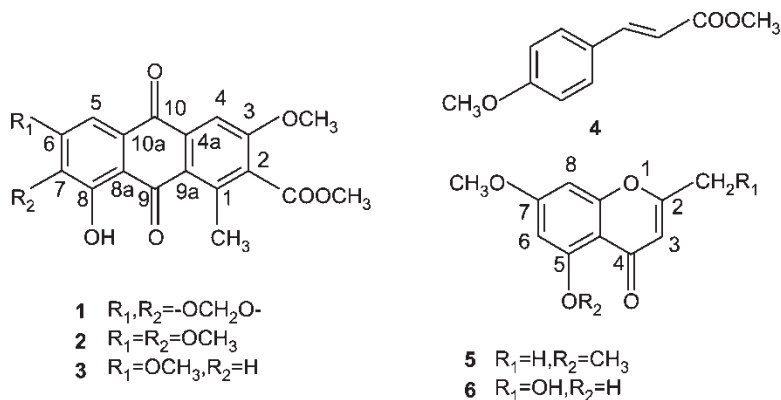


FIGURE 1 Structures of compounds 1–6.

out on a PROTÉGÉ 460 spectrometer and an ANTHELIE advanced spectrometer, respectively. TLC was performed on plates precoated with Merck RP-18 and silica gel GF₂₅₄, and HRTLC on silica gel H (5–7 μ). Separation and purification were performed by column chromatography on silica gel (160–200 and 200–300 mesh) and Merck RP-18 gel (40–63 μ).

Plant Material

The dried subterranean corms of *G. gandavensis* Van Houtt were collected in Chengdu, China in October, 2000, and identified by Professor Zuocheng Zao at the Chengdu Institute of Biology of the Chinese Academy of Sciences. A voucher specimen (No.w-2000108) is deposited at the Chengdu Institute of Biology, Chinese Academy of Sciences, China.

Extraction and Isolation

Powdered plant material (30 kg, 20–30 mesh) was extracted with EtOH (3 \times 100 L) at room temperature. After evaporation of EtOH under reduced pressure, 2.5 kg of viscous residue was obtained. This residue was suspended in H₂O and partitioned successively with light petroleum, CHCl₃, EtOAc and n-BuOH to give corresponding extracts of 325 g, 105 g, 250 g and 680 g. The light petroleum extract (325 g) was chromatographed over silica gel (gradiently eluted with light petroleum-acetone from 50:1 to 20:1) to give fractions A (10.5 g) and B (15.0 g). Fraction A (10.5 g) was subjected to silica gel column chromatography eluted with light petroleum-EtOAc (35:1) to yield a fraction (145 mg) that was rechromatographed on silica gel with cyclohexane-EtOAc (11:1) to give compounds **4** (47 mg) and **5** (35 mg). Fraction B (15.0 g) was chromatographed over silica gel using CHCl₃ as solvent to yield a yellow residue (2.1 g) and a white residue (560 mg). The yellow residue (2.1 g) was then chromatographed over RP-18 gel using MeOH-H₂O (5:1) to afford compounds **2** (78 mg), **1** (25 mg) and **3** (105 mg). The white residue (560 mg) was rechromatographed over silica gel with cyclohexane-acetone (10:1) followed by crystallization from CHCl₃ to afford compound **6** (75 mg).

Gandavensin A (**1**), orange needles (CHCl₃), mp 215.5–217.0°C. IR ν_{\max} cm⁻¹ (KBr): 3371, 2920, 2851, 1738, 1644, 1579, 1565, 1503, 1451, 1372, 1321, 1278, 1227, 1181, 1098, 1077, 1032, 997, 942, 786, 610, 403. UV-Vis λ_{\max} (nm) (log ϵ) in MeOH: 213 (3.86), 287 (4.06), 319 (3.48), 403 (3.42). EIMS (m/z , rel. int. %): 370 (M⁺, 100), 353 (25), 339 (15), 323 (5), 309 (7), 295 (10), 280 (3), 267 (6), 253 (4), 239, 225, 197, 183, 176, 152, 139, 126, 105, 91, 77, 69, 55; HR-EIMS m/z 370.0678 (calcd. for C₁₉H₁₄O₈, 370.0689); for ¹H NMR and ¹³C NMR data see Table I.

TABLE I NMR data of gandavensin A in CDCl₃ (400 MHz for ¹H, 100 MHz for ¹³C, δ in ppm)*

Position	C atom	δ _H (J in Hz)	δ _C	HMBC
1	C		141.0	
2	C		130.3	
3	C		158.9	
4	CH	7.57 (s)	107.4	C-10, C-9a, C-2, C-4a, C-3
4a	C		136.4	
5	CH	7.33 (s)	104.3	C-10, C-8a, C-7, C-6
6	C		153.2	
7	C		139.0	
8	C		145.3	
8a	C		115.3	
9	C		188.8	
9a	C		122.2	
10	C		182.0	
10a	C		128.1	
11	C		167.8	
1-CH ₃	CH ₃	2.74 (s)	19.4	C-1, C-2, C-9a
2-COOCH ₃	CH ₃	3.89 (s)	53.5	C-11
3-OCH ₃	CH ₃	3.96 (s)	57.5	C-3
6,7-OCH ₂ O	CH ₂	6.27 (s)	102.2	C-6, C-7
8-OH		12.54 (s)		C-8, C-7, C-8a

* Assignments were based on ¹H-¹H COSY, DEPT, HMQC and HMBC experiments.

Gandavensin B (**2**), orange needles (CHCl₃), mp 226.0–227.5°C. IR ν_{max} (cm⁻¹) (KBr): 3459, 3004, 2951, 1740, 1668, 1627, 1577, 1493, 1453, 1418, 1369, 1315, 1271, 1174, 1133, 1076, 971, 797, 678. UV-Vis λ_{max} (nm) (log ε) in MeOH: 204 (4.04), 283 (4.07), 316 (3.53), 418 (3.89). EIMS (*m/z*, rel. int. %): 386 (M⁺, 100), 371(43), 355 (18), 339 (11), 325 (7), 314 (3), 297 (4), 282 (2), 240 (3), 177 (4), 105 (2), 84 (3), 69 (5), 55 (4). HR-EIMS *m/z* 386.1048 (calcd. for C₂₀H₁₈O₈, 386.1002); for ¹H NMR and ¹³C NMR data see Table II.

Methyl 8-hydroxy-3,6-dimethoxy-1-methylanthraquinone-2-carboxylate (**3**), yellow needles (CHCl₃), mp 200.0–202.0°C. IR ν_{max} (cm⁻¹) (KBr): 3438, 2927, 2852, 1735,

TABLE II NMR data of gandavensin B in CDCl₃ (400 MHz for ¹H, 100 MHz for ¹³C, δ in ppm)*

Position	C atom	δ _H (J in Hz)	δ _C	HMBC
1	C		142.0	
2	C		131.1	
3	C		159.6	
4	CH	7.70 (s)	107.6	C-10, C-9a, C-2, C-4a, C-3
4a	C		137.3	
5	CH	7.37 (s)	103.2	C-10, C-8a, C-7, C-6, C-10a
6	C		157.5	
7	C		141.3	
8	C		156.7	
8a	C		113.1	
9	C		189.0	
9a	C		124.7	
10	C		181.7	
10a	C		128.1	
11	C		167.5	
1-CH ₃	CH ₃	2.72 (s)	19.9	C-1, C-2, C-9a
2-COOCH ₃	CH ₃	3.98 (s)	52.7	C-11
3-OCH ₃	CH ₃	4.03 (s)	56.5	C-3
6-OCH ₃	CH ₃	4.01 (s)	56.4	C-6
7-OCH ₃	CH ₃	4.02 (s)	60.9	C-7
8-OH		13.08		C-8, C-7, C-8a

* Assignments were based on ¹H-¹H COSY, DEPT, HMQC and HMBC experiments.

1629, 1579, 1512, 1454, 1389, 1321, 1249, 1118, 1066, 1005. UV-Vis λ_{\max} (nm) (log ϵ) in MeOH: 203 (3.94), 221 (3.95), 283 (4.00), 344 (3.14), 426 (3.28). EIMS (m/z , rel. int. %): 356 (M^+ , 70), 341(79), 325 (20), 309 (13), 295 (19), 267 (15), 162 (5), 139 (4), 84 (41), 71 (7), 69 (25), 56 (53), 41 (44). HR-EIMS m/z 356.0887 (calcd. for $C_{19}H_{16}O_7$, 356.0896). 1H NMR (400 MHz, $CDCl_3$) δ ppm: 6.68 (1H, d, $J = 2.8$ Hz, 7-H), 7.27 (1H, d, $J = 2.8$ Hz, 5-H), 7.71 (1H, s, 4-H), 3.92 (3H, s, 6-OCH₃), 3.98 (3H, s, 2-COOCH₃), 4.01 (3H, s, 3-OCH₃), 2.72 (3H, s, 1-CH₃), 12.13 (s, 8-OH). ^{13}C NMR (100 MHz, $CDCl_3$) δ ppm: 141.2 (1-C), 131.3 (2-C), 159.4 (3-C), 107.6 (4-C), 137.3 (4a-C), 107.4 (5-C), 165.4 (6-C), 106.8 (7-C), 165.2 (8-C), 111.4 (8a-C), 188.3 (9-C), 124.8 (9a-C), 182.3 (10-C), 134.0 (10a-C), 167.6 (2-COOCH₃), 56.5 (3-OCH₃), 55.9 (6-OCH₃), 52.7 (2-COOCH₃) and 19.7 (1-CH₃).

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References

- [1] Anonymous (1985), *The Annals of Chinese Plants (Zhongguo Zhiwuzhi)* (Science Press, Beijing) Vol. 16 (1), pp. 124–125.
- [2] Ali, A.A., Abdallah, O.M. and Steglich, W. (1989), *Phytochemistry* **28**, 281–282.
- [3] Viladomat, F., Codina, C., Llabres, J.M. and Bastida, J. (1986), *Int. J. Crude Drug Res.* **24**, 123–130.
- [4] Seilleur, P. (1977), *Bull. Rech. Agron. Gemblous* **12**, 121–134.
- [5] Salehian, A. (1973), *Bull. Trav. Soc. Pharm. Lyon* **17**, 86–90.
- [6] Jiangsu New Medical College (2000), *Dictionary of Chinese Herb Medicine* (Shanghai Science and Technology Press, Shanghai) Vol. 1, p. 2341.
- [7] Cameron, D.W., Deutscher, D.J., Feutrill, G.I. and Griffiths, P.G. (1981), *Aust. J. Chem.* **34**, 2401–2421.
- [8] Wat, C.-K., Towers, G.H.N. and Eberhardt, G. (1956), *J. Am. Chem. Soc.* **78**, 2832–2835.
- [9] Taylor, D.R., Warner, J.A. and Wright, J.A. (1977), *J. Chem. Soc., Perkin Trans. 1*, 397–405.
- [10] Kobayashi, M., Tawara, T., Tsuchida, T. and Mitsuhashi, H. (1990), *Chem. Pharm. Bull.* **38**, 3169–3171.
- [11] Huang, L. and Yu, D.Q. (2000), *Application of UV Spectrum in Organic Chemistry* (Science Press, Beijing) Vol. 2, p. 577.
- [12] Rauwald, H.W. and Miething, H. (1985), *Deut. Apoth. Ztg.* **125**, 101–103.
- [13] Yagi, A., Makino, K. and Nishioka, I. (1974), *Chem. Pharm. Bull.* **22**, 1159–1166.
- [14] Komura, H., Mizukawa, K., Minakata, H., Huang, H., Qin, G. and Xu, R. (1983), *Chem. Pharm. Bull.* **31**, 4206–4208.
- [15] Oshio, H., Naruse, Y. and Tsukui, M. (1978), *Chem. Pharm. Bull.* **26**, 2458–2464.
- [16] Gao, W.Y., Li, Y.M. and Zhu, D.Y. (1999), *Acta Bot. Sin.* **41**, 531–533.
- [17] Mammo, W., Dagne, E. and Steglich, W. (1992), *Phytochemistry* **31**, 3577–3581.